Multiproxy evidence of the Neoglacial expansion of Atlantic Water to eastern Svalbard

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Abstract. The main goal of this study is to reconstruct the paleoceanographic development of Storfjorden during the Neoglacial (∼4 cal ka BP). Storfjorden is one of the most important brine factories in the European Arctic and is responsible for deepwater production. Moreover, it is a climate-sensitive area influenced by two contrasting water masses: warm and saline Atlantic Water (AW) and cold and fresh Arctic Water (ArW). Herein, a multiproxy approach was applied to provide evidence for existing interactions between the inflow of AW and sea ice coverage, which are the major drivers of environmental changes in Storfjorden. The sedimentary and microfossil records indicate that a major reorganization of oceanographic conditions in Storfjorden occurred at ∼2.7 cal ka BP. The cold conditions and the less pronounced presence of AW in Storfjorden during the early phase of the Neoglacial were the prerequisite conditions for the formation of extensive sea ice cover. The period after ∼2.7 cal ka BP was characterized by alternating short-term cooling and warming intervals. Warming was associated with pulsed inflows of AW and sea ice melting that stimulated phytoplankton blooms and organic matter supply to the bottom. The cold phases were characterized by heavy and densely packed sea ice, resulting in decreased productivity. The ancient environmental DNA (aDNA) records of foraminifera and diatoms support the occurrence of the major pulses of AW (∼2.3 and ∼1.7 cal ka BP) and the variations in sea ice cover. The episodes of enhanced AW inflow were marked by an increase in the percentage of DNA sequences of monothalamous foraminifera associated with the presence of fresh phytodetritus. Cold and less productive intervals were marked by an increased proportion of monothalamous taxa known only from environmental sequencing.

1 Introduction

The northward flow of Atlantic Water (AW) is one of the major contributors of heat to the Arctic Ocean (Polyakov et al., 2017). Recent oceanographic data indicate a warming trend due to an increased inflow of AW towards the Arctic Ocean (Rudels et al., 2015, Polyakov et al., 2017). AW has been present along the western margin of Svalbard for at least the last 12 000 years (e.g., Werner et al., 2011; Rasmussen et al., 2014). One of the major intrusions of AW occurred during the early Holocene (10.8–6.8 cal ka BP). A distinct cooling and freshening of the west Spitsbergen shelf bottom-water masses occurred during the middle to late Holocene (6.8–1 cal ka BP) and were accompanied by glacier readvances in Svalbard, leading to the present-day conditions (Ślubowska-Woldengen et al., 2007; Telesiński et al., 2018). The paleoceanographic conditions in the Svalbard margins correlate closely with the sea surface temperature (SST) variations in the Nordic Seas and confirm that the Svalbard area is highly sensitive to fluctuations in the inflow of AW (Ślubowska-Woldengen et al., 2007; Werner et al., 2013). Conversely, until the 1990s eastern Svalbard was recognized as an area exclusively influenced by the East Spitsbergen Current (ESC), which carries cold, less saline
The aim of this study is to reconstruct the paleocceanographic development of Storfjorden during the Neoglacial at multicentennial resolution. We assumed that the periodic intensification of the AW inflow to the west Spitsbergen shelf during the Neoglacial also resulted in the appearance of AW in eastern Spitsbergen, similar to the conditions in the early Holocene (e.g., Łącka et al., 2015), affecting the density and extent of sea ice cover in the area. A multiproxy approach comprising sedimentary, microfossil and molecular records was applied to provide evidence for the interactions between the inflow of AW and sea ice coverage in Storfjorden. The ancient environmental DNA (aDNA) analysis targeted diatoms and nonfossilized monothalamous foraminifera. Both of these groups are hardly preserved in fossil records from the Svalbard fjords (Pawłowska et al., 2014) and shelf areas (Zimmermann et al., 2019, and references therein). Recent studies have demonstrated that the analysis of genetic material obtained directly from environmental samples (so-called environmental DNA) is an efficient method for performing biodiversity surveys across time and space (Thomsen and Willerslev, 2015). The content of environmental DNA samples may be analyzed by DNA metabarcoding, which consists of high-throughput sequencing of taxonomically informative DNA fragments called metabarcodes. The identification of short, species-specific DNA fragments (so-called “barcodes”) allows us to obtain species-level assignments of modern and ancient DNA sequences (Herbert et al., 2003). The further demonstration that DNA can be preserved in the environment across geological timescales opened new avenues for paleoclimatic and paleoceanographic studies. Recent studies have demonstrated the preservation of DNA in marine sediments for tens to hundreds of thousands of years. An aDNA approach was successfully applied to trace the Holocene history of dinoflagellates, haptophytes (e.g., Coolen et al., 2009, 2013; Boere et al., 2009) and foraminifera in deep-sea (Lejzerowicz et al., 2013) and coastal areas (Pawłowska et al., 2014, 2016). The study of Pawłowska et al. (2016) was the first attempt to utilize foraminiferal aDNA as a paleoenvironmental proxy. This study supported the existence of extremely diverse foraminiferal assemblages. The richness of the foraminiferal community revealed by the molecular record was much higher than that in the fossil record (Pawłowska et al., 2014), mainly due to the detection of nonfossilized monothalamous taxa. The molecular data correlated well with environmental changes and revealed even small changes that were not clearly indicated by other proxy records. The combination of aDNA studies with the analysis of microfossils and sedimentary proxies provides a powerful means to reconstruct past environments more comprehensively.

2 Study area

Storfjorden is located in southeastern Svalbard between the islands of Spitsbergen, Edgeøya and Barentsøya (Fig. 1).
Storfjorden is ∼190 km long and its main basin is ∼190 m deep. Two narrow and shallow passages (Heleysundet and Freemansundet) connect northern Storfjorden to the Barents Sea. To the south, a 120 m deep sill separates the main basin from Storfjordrenna. Storfjordrenna is 245 m long, with a depth varying from 150 to 420 m.

The water masses in Storfjorden are composed primarily of exogenous Atlantic and Arctic waters as well as mixed waters that have formed locally. The Atlantic Water is transported northwards by the Norwegian Atlantic Current (NwAC), which branches off in the Barents Sea into the West Spitsbergen Current and North Cape Current (Loeng, 1991; Blindheim and Østerhus, 2005) (Fig. 1a). In addition to AW, cold and less saline Arctic Water (ArW) is transported to the Barents Sea via the East Spitsbergen Current and Bear Island Current (Hopkins, 1991) (Fig. 1a). Arctic Water (ArW) from the Arctic Ocean and the Barents Sea enters Storfjorden via two passages to the northeast and continues along the inner shelf of Svalbard as a coastal current (Fig. 1b). AW is characterized by temperatures > 3 °C and salinity > 34.95, while the temperature and salinity of ArW are < 0 °C and 34.3–34.8, respectively. The presence of locally formed water masses is a result of the interactions between AW, ArW and meltwater. Skogseth et al. (2005) listed six local water masses: meltwater (MW), polar front water (PW), east Spitsbergen water (ESW), brine-enriched shelf water (BSW), Storfjorden surface water (SSW) and modified Atlantic Water (MAW). BSW is formed due to the release of large amounts of brine during polynya events and the intensive formation of sea ice (Haarpainter et al., 2001; Skogseth et al., 2004, 2005) and is characterized by salinities exceeding 34.8 and temperatures below −1.5 °C (Skogseth et al., 2005).

The sedimentary environment in Storfjorden is classified as a low-energy, high-accumulation environment, which is characteristic of inner fjords. The area is sheltered from along-shelf bottom currents and is affected by high terrigenous inputs; therefore, deposition prevails over sediment removal by bottom currents (Winklemann and Knies, 2005). The primary productivity is high and strongly depends on the sea ice formation as well as the seasonal duration of the marginal ice zone (Winkelman and Knies, 2005).

3 Materials and methods

3.1 Marine sediment core

The 55 cm long sediment core ST_1.5 was taken with a gravity corer in Storfjorden retrieved with the R/V Oceania in August 2014. The sampling station was located at 76°53.181' N and 19°27.559' E at a depth of 153 m (Fig. 1). The salinity and temperature of the water column at the coring station were measured with a mini-CTD Sensordata SD204 at intervals of 1 s (Fig. 2). The core was stored at 4 °C and shipped to the Institute of Oceanology PAS for further analyses.

In the laboratory, the core was extruded and cut into 1 cm slices. During cutting, sterile subsamples for ancient DNA (aDNA) analyses were taken at 4 cm intervals. To avoid external and/or cross-contamination the thin layers of sediment that were in contact with underlying or overlying sediments were removed using a sterile spatula. Samples for aDNA analyses were kept frozen at −20 °C. Samples for other proxy analyses were taken every 2 cm.
Table 1. Raw and calibrated AMS$^{14}$C dates used in the age model. B stands for bivalve shells, while F stands for benthic foraminifera tests.

<table>
<thead>
<tr>
<th>Core depth (cm)</th>
<th>Material</th>
<th>Raw AMS$^{14}$C</th>
<th>Cal. years BP ±2σ</th>
<th>Cal. years BP used in age model</th>
</tr>
</thead>
<tbody>
<tr>
<td>2.5</td>
<td>Nuculana pernula (B)</td>
<td>107.38 ± 0.33 pMC</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>5.5</td>
<td>Yoldiella lenticula (B)</td>
<td>290 ± 30 BP</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>14.5</td>
<td>Turritella erosa (B)</td>
<td>2020 ± 30 BP</td>
<td>1356–1555</td>
<td>1500</td>
</tr>
<tr>
<td>43.5</td>
<td>Yoldiella solituda (B)</td>
<td>3010 ± 50 BP</td>
<td>2484–2787</td>
<td>2700</td>
</tr>
<tr>
<td>46.5</td>
<td>Nonionellina labradorica (F)</td>
<td>4490 ± 40 BP</td>
<td>4400–4701</td>
<td>4500</td>
</tr>
<tr>
<td>52.5</td>
<td>Yoldiella lenticula (B)</td>
<td>7545 ± 35 BP</td>
<td>7803–7989</td>
<td>7890</td>
</tr>
</tbody>
</table>

3.3 Grain size analysis

The samples for grain size analyses were freeze-dried and milled. The measurements were performed using a Mastersizer 2000 particle laser analyzer coupled to a Hydro MU device (Malvern, UK). The samples were treated with ultrasound to avoid aggregation. The raw data were analyzed using GRADISTAT v.8.0 software (Blott and Pye, 2001). The mean 0–63 µm grain size [ϕ] was calculated via the logarithmic method of moments. The sediment fraction > 500 µm was used to reconstruct an ice-rafted debris (IRD) record. The grains were counted under a stereomicroscope, and the amount of IRD is reported as the concentration (i.e., the number of grains per gram of dry sediment) [grains g$^{-1}$] and the flux [grains cm$^{-2}$ yr$^{-1}$].

3.4 Benthic foraminifera assemblages

Prior to the analysis of benthic foraminifera, samples were wet-sieved through meshes with 500 and 100 µm openings and dried at 60 °C. Samples with large quantities of tests were divided using a microsplitter. At least 300 specimens of benthic foraminifera were isolated from each sample and collected on micropaleontological slides. Benthic foraminifera specimens were counted and identified to the lowest possible taxonomic level. The quantity of foraminifera is presented as the concentration (i.e., the number of individuals per gram of dry sediment) [ind. g$^{-1}$] and the flux [ind. cm$^{-2}$ yr$^{-1}$]. Foraminifera species were grouped according to their ecological tolerances. Four groups of indicators were distinguished: AW and/or frontal zone indicators, ArW indicators, bottom current indicators, and glaciomarine species (Majewski et al., 2009). The morphologically similar species Islandiella norcrossi and Islandiella helenae are reported as Islandiella spp.

3.5 Stable isotope analysis

Carbon and oxygen stable isotope analyses were performed on Cibicidoides lobatulus tests selected from 27 sediment layers. Approximately 10 to 12 specimens were collected from each sample and subjected to ultrasonic cleaning. The measurements were performed on a Finnigan MAT 253 mass spectrometer coupled to a Kiel IV carbonate preparation de-
vice at the University of Florida. The resulting values are expressed in standard δ notation relative to Vienna Pee Dee Belemnite (VPDB).

### 3.6 Ancient DNA analysis

The total DNA was extracted from approximately 10 g of sediment using a Power Max Soil DNA extraction kit (MoBio). The foraminiferal SSU rDNA fragments containing the 3′ hypervariable region were PCR-amplified (PCR: polymerase chain reaction) using primers tagged with unique sequences of five nucleotides appended to their 5′ ends (denoted by Xs), namely, the foraminifera-specific forward primer s14F1 (5′-XXXXXCGGACACACTGAGGATTGACAG-3′) and the reverse primer s15 (5′-XXXXXXXXCTCTACATAATCATGAAAG-3′). The diatom DNA fragment located in the V4 region was amplified with the forward DIV4for (5′-XXXXXXXXCGCGGTAAAATCCAGCTCCATAG-3′) and reverse DIV4rev3 (5′-XXXXXXXXCTCTGACAATGGAATACGAATA-3′) primers tagged with a unique combination of eight nucleotides (denoted by Xs) attached at each primer’s 5′ end. The amplicons were purified using the High Pure PCR Cleanup Micro Kit (Roche) and quantified using a Qubit 2.0 fluorometer. Samples were pooled in equimolar quantities, purified using the High Pure PCR Cleanup Micro Kit (Roche) and quantified using a Qubit 2.0 fluorometer. Samples were then diluted to a concentration of 10 ng μl−1 and analyzed on a 2% agarose gel. Each sample was run in triplicate, and the experiment was repeated at least twice.

### 3.7 Sediment grain size

The sediment was classified as medium to coarse silt throughout the core. The sediment accumulation rate (SAR) prior to ∼2.7 cal ka BP was 0.002 cm yr−1. The approximately 10-fold increase in SAR was noted at ∼2.7 cal ka BP, when it increased to 0.023 cm yr−1. During the last 1.5 cal ka BP, SAR decreased to 0.01 cm yr−1 (Fig. 4). The amount of IRD was the highest prior to ∼2.7 cal ka BP, reaching up to 83 grains cm−2 yr−1. After ∼2.7 cal ka BP, the amount of IRD was relatively stable and did not exceed 18 grains cm−2 yr−1. The IRD flux decreased slightly over time to 0.37 grains cm−2 yr−1, except for one peak reaching 0.8 grains cm−2 yr−1 at ∼2.6 cal ka BP (Fig. 4).

The mean grain size of the 0–63 µm fraction had its highest value (5.8 µm) at ∼2.7 cal ka BP (Fig. 4). After ∼2.4 cal ka BP a slight but continuous reduction in the mean 0–63 µm grain size was noted. The minimum grain size (6.23 µm) was recorded at the top of the core (Fig. 4).

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**Figure 3.** Age–depth model of the ST_1.5 core. The grey silhouettes show the probability distribution of calendar dates that were obtained by calibration of the individual 14C dates used for the age model. The dotted line shows the age–depth model derived from a linear interpolation between the dates.

**Figure 4.** Sediment depth in cm above 0 cm for the top 50 cm of the core. The 50% probability distribution of calendar dates is shown for each sample. The blue line shows the age–depth model derived from a linear interpolation between the dates.
Sedimentological and micropaleontological data plotted versus age. The sediment accumulation rate (SAR), mean grain size of the 0–63 µm fraction, ice-rafted debris (IRD) flux and number of grains per gram of sediment, oxygen ($\delta^{18}O$) and carbon ($\delta^{13}C$) stable isotopes in benthic foraminiferal tests, and the flux and abundance of foraminifera are presented.

4.3 Stable isotopes

The two $\delta^{18}O$ data points prior to $\sim 2.7$ cal ka BP recorded values of 3.55 ‰ and 3.69 ‰ vs. VPDB. Between $\sim 2.7$ and $\sim 1.5$ cal ka BP, $\delta^{18}O$ showed the strongest variation, with values ranging from 3.28 ‰ to 3.77 ‰ vs. VPDB. After $\sim 1.5$ cal ka BP, $\delta^{18}O$ became slightly lighter (3.43 ‰–3.64 ‰ vs. VPDB), except for one peak noted in the uppermost layer of the core, where $\delta^{18}O$ reached 3.87 ‰ vs. VPDB (Fig. 4).

In the period prior to $\sim 2.7$ cal ka BP, recorded values of $\delta^{13}C$ reached 0.92 ‰ and 1.12 ‰ vs. VPDB. Slightly heavier

$\delta^{13}C$ (up to 1.46 ‰ vs. VPDB) was observed between $\sim 2.7$ and $\sim 1.5$ cal ka BP. The gradual decrease was recorded from $\sim 1.5$ cal ka BP to the present, reaching 0.81 ‰ vs. VPDB at the top of the core (Fig. 4).

4.4 Benthic foraminifera assemblages

A total of 8647 fossil foraminifera specimens belonging to 47 species were identified (Supplement Fig. S1). The number of foraminifera individuals in a sample varied from 156 to 2610 ind. g$^{-1}$, and the lowest abundances were observed prior to $\sim 2.7$ cal ka BP (Fig. 4). A short-term decrease in foraminifera abundance was observed between 2.2 and 1.7 cal ka BP, with values reaching as low as 304 ind. g$^{-1}$. The abundance maxima were noted at 2.3, 1.5 and 0.6 ka BP, with values reaching 2524, 2584 and 2610 ind. g$^{-1}$, respectively. The foraminiferal flux was low and relatively stable throughout the core, with values that did not exceed 1 ind. cm$^{-2}$ yr$^{-1}$, except for two peaks at 2.3 and 1.5 ka BP, when the flux reached 2.2 ind. cm$^{-2}$ yr$^{-1}$ for both peaks (Fig. 4).

The most abundant species was *Cassidulina reniforme*, with densities reaching up to 900 ind. g$^{-1}$. The other species that constituted the majority of the foraminiferal assemblage were *Buccella frigida*, *Cibicidoides lobatulus*, *Elphidium excavatum*, *Islandiella* spp., *Melonis barleeanum* and *Nonionellina labradorica*. The abundances of the dominant species followed a general trend, with maxima at $\sim 2.3$ cal ka BP and after $\sim 1.7$ cal ka BP and minima prior to $\sim 2.7$ cal ka BP and between 2.3 and 1.7 cal ka BP (Fig. 5). The foraminiferal assemblage prior to $\sim 2.7$ cal ka BP was dominated by indicators of AW inflow and/or frontal zones and glaciomarine taxa (Fig. 5). The most abundant species were *Nonionellina labradorica* and *Melonis barleeanum*, as well as *Cassidulina reniforme* and *Elphidium excavatum*, which together accounted for up to 60 % of the foraminiferal abundance (Fig. 5). After $\sim 2.7$ cal ka BP, there were AW and/or frontal zone indicator peaks recorded at 2.4 and 1.8 cal ka BP, when the percentages increased to 33 % and 28 % of the total abundance, respectively. The period between $\sim 2.4$ and $\sim 1.8$ cal ka BP was characterized by an increase in the percentage of sea ice indicators (*B. frigida* and *Islandiella* spp.), which accounted for up to 25 % of the total foraminiferal abundance. Additionally, a short-term peak in the glaciomarine taxa, reaching up to 49 % of the foraminiferal assemblage, was recorded between 2.5 and 2.1 cal ka BP. Also, the overall maximum of glaciomarine species abundance was recorded between 1.7 and 0.5 cal ka BP, ranging from 25 % to 43 % of the foraminiferal assemblage (Fig. 5). A decrease in the relative abundance of glaciomarine species was observed after $\sim 0.5$ cal ka BP and was followed by an increase in the AW and/or frontal zone indicators and a single peak in the percentage of bottom current indicators, which reached 42 % and 19 %, respectively (Fig. 5).
4.5 Foraminiferal aDNA sequences

A total of 1,499,889 foraminiferal DNA sequences were clustered into 263 OTUs, and 20 remained unassigned. The remaining OTUs were assigned to Globigerinida (5 OTUs), Robertinida (1 OTU), Rotaliida (49 OTUs), Textulariida (18 OTUs), Monothalamea (163 OTUs) and Milolida (7 OTUs). The majority of sequences belonged to Monothalamea (60%) and Rotaliida (31%) (Fig. S2). Herein, we focus on Monothalamea, which is the dominant component of the foraminiferal aDNA record.

The most important components of the monothalamous assemblage were Micrometula sp., Cylindrogullmia sp., Hippocrepinella hirudinea, Ovammina sp., Nemogullmia sp., Tinogullmia sp., Cedhagenia saltatus, undetermined allogromiids belonging to clades A and Y (herein called “allogromiids”), and sequences belonging to taxa known exclusively from environmental sequencing (herein called “environmental clades”). Herein, the term “allogromiid” refers to monothalamous foraminifera with organic or predominantly organic test walls (Gooday, 2002). Morphological and molecular evidence indicates that “allogromiids” are not a coherent taxonomic group but are scattered between several monothalamous clades (Pawlowski et al., 2002). “Clade” refers to phylogenetic clades defined by molecular data. The clade is traditionally defined as a group of organisms that includes a common ancestor and all the descendants. The sequences belonging to allogromiids were present throughout the core, accounting for 16%–31.7% of all the foraminiferal sequences. The exceptions were the intervals from ~4.0 to 2.4 cal ka BP and ~1.7 cal ka BP, when the contribution of allogromiid sequences decreased to less than 10% (Fig. 6). The majority of the allogromiids belonged to clade Y, which made up to 100% of the allogromiid sequences. Only at 1.6–1.7 and 2.4–2.6 cal ka BP did most of the allogromiid sequences belong to clade A. Additionally, allogromiids belonging to clade I were noted at ~2.4 cal ka BP, when they made up 0.88% of allogromiid sequences (Fig. 7).

The periods prior to ~2.4 and ~1.7 cal ka BP were marked by the disappearance of sequences belonging to C. saltatus, Nemogullmia sp. and the environmental clades, followed by an increase in the percentages of sequences belonging to Micrometula sp., Ovammina sp., Tinogullmia sp., Shepheardella sp. and Cylindrogullmia sp. (Fig. 6).

4.6 Diatom aDNA sequences

A total of 824,697 diatom DNA sequences were clustered into 221 OTUs (Fig. S3). The most abundantly sequenced diatom taxa were Thalassiosira spp, which made up 61.1% of diatom sequences. Other abundantly sequenced taxa were Chaetoceros sp. and T. antarctica, which made up 8.5% and 11.5% of sequences, respectively. The sequences of Thalassiosira sp. were most abundant between ~2.2 and ~1.9 cal ka BP, accounting for up to 85% of all diatom sequences. The lowest percentage (14%) of Thalassiosira sp. was recorded at ~0.4 cal ka BP. Sequences assigned to T. antarctica were recorded throughout the core, and their percentages were the highest at ~3.3 and ~2.6 cal ka BP, reaching up to 13% and 19%, respectively (Fig. 8). Sequences of T. hispida were also noted throughout the core and constituted 4.7% of diatom sequences in the uppermost layer. In the remaining samples, T. hispida sequences did not exceed 1%. The percentage of sequences of Chaetoceros sp.
The dominant components of the monothalamous assemblages. The abundance is expressed as the percentage of the monothalamous sequences, and the most abundantly sequenced taxa are presented. The trend (two-point average) is indicated with a dashed line.

Figure 6. The percentage share of certain clades in the allogromiid sequences.

Navicula sp. constituted an important part of the diatom assemblage at \( \approx 3.3 \) and \( \approx 1.9 \) cal ka BP, accounting for up to 25.5% and 10% of all diatom sequences, respectively. In the remaining samples, the abundance of Navicula sp. did not exceed 5% (Fig. 8).

5 Discussion

The ST_1.5 age model is based on the linear interpolation between the four AMS\(^{14}\)C dates; thus, the age control of the core should be treated with caution. However, the timing of major environmental changes revealed by the ST_1.5 multi-proxy record is in agreement with other records from the region (e.g., Sarnthein et al., 2003; Risebrobakken et al., 2010; Berben et al., 2017). Moreover, the major pulses of AW that were recorded \( \approx 2.3 \) and 1.7 cal ka BP correlated well with winter and summer SST maxima recorded in the 23258-2 core (Sarnthein et al., 2003).

5.1 The period from 4 to 2.7 cal ka BP

Prior to \( \approx 2.7 \) cal ka BP, the ST_1.5 sedimentary record displayed relatively higher IRD delivery and a relatively lower 0–63 µm sediment fraction than in the following period (Fig. 4). These results are in agreement with the record from Storfjordrenna (Łącka et al., 2015), where peaks in IRD were noted during the Neoglacial and were attributed to increased iceberg rafting due to fluctuations in the glacial fronts (e.g., Forwick et al., 2010). The coarser 0–63 µm fraction may suggest the winnowing of fine-grained sediment; however, foraminiferal fauna showed no clear response to sediment removal.

The foraminiferal flux and abundance prior to 2.7 cal ka BP reached their lowest values (Fig. 4). Previous studies reported a decrease in the concentration of benthic foraminifera in Storfjorden at that time, which was attributed to the presence of extensive ice cover (Rasmussen and Thomsen, 2015; Knies et al., 2017). The dominant
components of the ST_1.5 foraminiferal assemblage were *C. reniforme* and *M. barleeanum* (Fig. 5). The presence of *C. reniforme* and *M. barleeanum* is associated with cooled and salty AW (e.g., Hald and Steinsund, 1996; Jernas et al., 2013). Moreover, these species are also associated with the presence of phytodetritus, which may be related to the delivery of fresh organic matter observed in frontal zones and/or near the sea ice edge (Jennings et al., 2004). The presence of sea ice may also be indicated by the relatively light foraminiferal δ¹³C (Fig. 4), as well as the highest percentage of the sea ice species *Thalassiosira antarctica* (see Ikävalko, 2004; Fig. 8). However, the low sampling resolution during that period precluded us from making a general conclusion, and the latter assumptions should be confirmed by further studies.

5.2 The period from 2.7 to 0.5 cal ka BP: episodes of AW inflow at ∼ 2.3 and 1.7 cal ka BP

After ∼ 2.7 cal ka BP, the increase in SAR was followed by a decrease in the mean grain size of the 0–63 µm fraction and in the IRD delivery (Fig. 4). The 10-fold increase in SAR most likely resulted from the intensive supply of turbid meltwater from advancing glaciers and the consequent intensive sedimentation. Moreover, the accumulation of fine sediment may also be enhanced by the slowdown of the bottom currents, indicated by the finer 0–63 µm sediment fraction (Fig. 4). On the other hand, a decrease in IRD delivery may suggest that central Storfjorden was not impacted by iceberg rafting at that time. In contrast, Rasmussen and Thomsen (2015) suggested glacial advance, followed by intensive ice rafting and meltwater delivery to Storfjorden at that time. According to Knies et al. (2017), inner Storfjorden was covered by densely packed sea ice between ∼ 2.8 and 0.5 cal ka BP. Therefore, the decreasing IRD in the ST_1.5 core may result from the presence of a sea ice cover that reduced iceberg rafting, while the majority of coarse-grained material settled in the proximity to the glacial fronts. Similar conclusions have been stated by Forwick and Vorren (2009) and Forwick et al. (2010), who assumed that the enhanced formation of sea ice along the west Spitsbergen coast trapped icebergs inside the Isfjorden system.

The foraminiferal fauna in central Storfjorden revealed more than a 10-fold increase in flux and abundance, followed by short-term fluctuations after ∼ 2.7 cal ka BP (Fig. 4). The latter may suggest favorable conditions for foraminiferal growth. The major peaks in the total foraminiferal abundance (Fig. 4), followed by the peaks in the percentage of AW foraminiferal indicators (Fig. 5), were noted at ∼ 2.3 and ∼ 1.7 cal ka BP. These peaks were associated with the occurrence of sequences of *T. hispida* (Fig. 8), a diatom species characteristic of subpolar and temperate regions (Katsuki et al., 2009). The timing of the changes described above is in accordance with the findings of Sarntheim et al. (2003), who reported two intervals of a remarkably warmer sea surface on the western continental margin of the Barents Sea at ∼ 2.2 and ∼ 1.6 cal ka BP, which was attributed to short-term pulses of warm AW advection. Other records also indicated AW inflow to the western and northern Barents Sea as well as to the western Spitsbergen continental margin during the middle to late Holocene (e.g., Risebrobakken et al., 2010;
Müller et al., 2012; Groot et al., 2014; Berben et al., 2014; 2017). During the mid-Holocene, AW inflow to the Barents Sea was relatively stable. The environmental conditions became more unstable in the late Holocene, with periodic cooling of surface waters, the presence of AW and/or chilled AW near the bottom, and more extensive seasonal sea ice cover (Risebrobakken et al., 2010; Berben et al., 2014; Groot et al. 2014). The timing of these changes differed between the study settings: in the western Barents Sea, it was ~1.1 to 1.5 cal ka BP (Berben et al., 2014; Groot et al., 2014), while in the southwestern Barents Sea, the change in environmental conditions was recorded at ~2.5 cal ka BP (Risebrobakken et al., 2014). In contrast, the northern Barents Sea experienced surface water cooling and more extensive sea ice cover prior to 2.7 cal ka BP. The increasing influence of AW was observed after 2.7 cal ka BP (Berben et al., 2017). Our foraminiferal and diatom aDNA records confirm the presence of AW intrusions in Storfjorden after ~2.7 cal ka BP that may have caused an episodic breakup of sea ice cover and permitted primary production as well as the development of benthic biota, including foraminifers.

The pulses of AW inflow at 2.3 and 1.7 cal ka BP were marked by the maxima of the foraminiferal flux (Fig. 4) and by peaks in the abundance of species associated with highly productive environments, such as *M. barleeanum* and *N. labradorica* (Fig. 5). Moreover, the presence of diatom aDNA sequences throughout the core (Fig. 8) may suggest continuous primary production. However, the responses of the benthic foraminifera assemblage to the pulses of AW at ~2.3 and ~1.7 cal ka BP are slightly different. The dominant components of the foraminiferal assemblage at ~2.3 cal ka BP were *M. barleeanum* and *E. excavatum*, while at ~1.7 cal ka BP, *N. labradorica* and *C. reniforme* were dominant (Fig. 5). The major difference in environmental conditions between these two “AW episodes” was the coarser 0–63 µm sediment fraction noted at ~2.3 cal ka BP; this may indicate more intensive winnowing of fine sediment grains, which would have created favorable conditions for the development of opportunistic species, such as *E. excavatum*.

In contrast, the interval between 2.3 and 1.7 cal ka BP featured variable δ¹³C and δ¹⁸O, followed by a decrease in the foraminiferal flux and abundance (Fig. 4). The foraminiferal assemblage at this time was dominated by glaciomarine and sea ice taxa (Fig. 5), which indicate more severe environmental conditions with extensive sea ice cover and suppressed productivity.

The alternate cooling and warming periods described above were also reflected in the aDNA record of monothalamous foraminifera. During the periods with more severe environmental conditions (i.e., time intervals of 2.2–1.9 and 1.3–0.4 cal ka BP), the monothalamous foraminifera were dominated by allogromids belonging to clade Y (*Nemogullmia* sp., *C. saltatus*) and monothalamids belonging to so-called environmental clades (Fig. 6). A considerable portion of the allogromiid sequences in the ST_1.5 core belong to clade Y (Fig. 7), which is primarily composed of taxa known only from environmental sequencing that have previously been noted in modern sediments in the Spitsbergen fjords (Pawłowska et al., unpublished). Clade Y has also been abundantly sequenced in the coastal areas off Scotland, characterized by high levels of environmental disturbances (Pawłowski et al., 2014); this might suggest its high tolerance to environmental stress. *C. saltatus* was found by Gooday et al. (2011) in the Black Sea, and its occurrence in areas with high levels of pollution suggests that it is an opportunistic species with a high tolerance for environmental disturbances. In addition, so-called environmental clades are composed of monothalamous taxa known exclusively from environmental sequencing (Lecroq et al., 2011). The aforementioned taxa nearly disappeared during the episodes of enhanced AW inflow at ~2.4 and ~1.7 cal ka BP, and the monothalamous assemblage was dominated at that time by *Micrometula* sp., *Ovammina* sp., *Stepheardella* sp., *Tinogullinia* sp., *Cylindrogullmia* sp. and allogromids belonging to clade A (Figs. 6, 7). All these taxa have recently been observed in the fjords of Svalbard and Novaya Zemlya (e.g., Gooday, et al., 2005; Majewski et al., 2005; Sabbattini et al., 2007; Pawłowska et al., 2014; Korsun and Hald, 1998; Korsun et al., 1995). *Cylindrogullmia* and *Micrometula* are dependent on the presence of fresh phytodetritus (Alve, 2010). *Ovammina* sp. feeds on diatoms and other forms of microalgae (Goldstein and Alve, 2011). Similarly, the presence of *Tinogullinia* is largely controlled by the presence of organic material on the seafloor. High concentrations of *Tinogullinia* have been found in coastal (Cornelius and Gooday, 2004) and deep-sea regions (Gooday, 1993) within phytodetrital aggregates.

The taxa that dominated the monothalamous assemblage during warm intervals seem to be responsive to the delivery of organic matter and may flourish during phytoplankton blooms associated with the settling of organic matter (e.g., Alve, 2010; Sabbattini et al., 2012, 2013). The pulses of AW inflow may be associated with phytoplankton blooms stimulated by sea ice melting and with the organic matter supply to the bottom (see Łącka et al., 2019). The continuous aDNA record of the sea ice diatom *T. antarctica* (Fig. 8) suggests the presence of at least seasonal ice cover in the study area. On the other hand, the episodes of AW inflow were associated with the occurrence of the open-water taxa *T. hispida* (Fig. 8). The occurrence of sequences of both these taxa suggests the formation of ice cover during winter–spring, followed by ice-free summers. A similar scenario was proposed by Berben et al. (2017), who suggested increased AW to eastern Svalbard and partial summer sea ice occurrence after 2.7 cal ka BP. According to the record of Łącka et al. (2019) from Storfjordrenna, the sea ice melting induced the production of brine, which may launch convective mixing and nutrient resupply from the bottom; this stimulated primary production.
Conversely, the colder phases of the Neoglacial were characterized by heavy and densely packed sea ice, resulting in limited productivity (Knies et al., 2017). The presence of T. antarctica sequences and the disappearance of T. hispida (Fig. 8) may suggest that primary production was associated with sea ice. Furthermore, the monothalamous assemblage was less diverse and was dominated by more opportunistic taxa, which may indicate a reduced supply of organic matter to the bottom.

5.3 The period after 0.5 cal ka BP

Modern-like conditions were established in Storfjorden at ∼0.5 cal ka BP (Knies et al., 2017). The ST_1.5 record displayed a decrease in SAR compared to the preceding period, a decreasing 0–63 µm fraction and low IRD delivery (Fig. 4), which may indicate reduced glacial impact. Moreover, the peak of heavy δ18O recorded on the core top (Fig. 4) may suggest the presence of AW or slightly increased salinity. Similarly, Berben et al. (2014) recorded δ18O values that suggested a minor increase in salinity, while foraminiferal fauna showed slightly lower salinities in the western Barents Sea at that time. The latter is in accordance with records from the Fram Strait (e.g., Werner et al., 2013) and the western Spitsbergen shelf (Cabedo-Sanz and Belt, 2016), which suggest episodes of freshening of the surface water masses associated with alternating sea ice increases and ice-free conditions in the late Holocene. Additionally, the records of Rasmussen and Thomsen (2014) and Knies et al. (2017) from Storfjorden indicated seasonally variable sea ice cover. Moreover, the majority of diatom aDNA sequences found in the ST_1.5 record after ∼0.5 cal ka BP belonged to Chaetoceros sp. (Fig. 8), a taxa that is observed in surface waters and is almost entirely absent under sea ice (Rožańska et al., 2008). High abundances of Chaetoceros are often associated with highly productive surface waters (Cremer, 1999). Riquel–Hernández et al. (2017) also noted an increased abundance of Chaetoceros sp. and enhanced algal productivity in Storfjorden after 2.0 cal ka BP, which was associated with the vicinity of the Arctic Front. However, the aDNA record of the monothalamous foraminifera at ∼0.4 cal ka BP displayed relatively high percentages of taxa that dominated during the colder intervals of the Neoglacial (Fig. 6). This may be related to the recovery from the Little Ice Age (LIA) and consequently from the temporarily deteriorated environmental conditions (D’Andrea et al., 2012). However, due to the low resolution during the LIA, a detailed interpretation is not possible. Therefore, further studies are required to confirm the latter conclusion.

5.4 Paleoceanographic implications

Our record revealed a two-phase Neoglacial, with a major shift in environmental conditions at ∼2.7 cal ka BP. According to the ST_1.5 proxy records, the Neoglacial in Storfjorden was not a constantly cold period, but comprised alternating short-term cooling and warming periods, associated with variability in sea ice coverage and productivity. The Neoglacial cooling was documented in various proxy reconstructions from the Nordic Seas (e.g., Jennings et al., 2002; Moros et al., 2004; Consolaro et al., 2018). However, there is growing evidence of shifts in environmental conditions in the Nordic Seas region in the Neoglacial, whose timings are in accordance with our record. An alkenone record from the Norwegian Sea revealed a significant drop in sea surface temperature at 2.7 cal ka BP (Calvo et al., 2002). Risebrobakken et al. (2010) recorded a change in oceanographic conditions in the SW Barents Sea at ca. 2.5 cal ka BP. The episodes of reduced surface and subsurface salinity were recorded after 2.5 cal ka BP, which was attributed to the expansion of coastal waters and the occurrence of more sea ice (Risebrobakken et al., 2010). Berben et al. (2017) recorded a shift at ∼2.7 cal ka BP from the marginal ice zone to Arctic frontal conditions in the eastern Barents Sea. They observed a continuous cooling trend from ∼5.9 to 2.7 cal ka BP, with increased seasonal sea ice with less open-water conditions, lower temperatures, and decreased AW influence, whereas after 2.7 cal ka BP the influence of AW was variable but generally generally increasing. The period was characterized by low insolation associated with surface cooling and enhanced formation of sea ice with reduced sea ice melt (Berben et al., 2017).

Moreover, our evidence of the presence of AW in Storfjorden during the Neoglacial supports previous suggestions that AW inflow during the late Holocene was strong enough to also reach the eastern coasts of Svalbard (e.g., Łącka et al., 2015). Episodic increases in the AW during the late Holocene were also observed in the northern Barents Sea (Duplessy et al., 2001; Lubinski et al., 2001), the eastern Barents Sea (Berben et al., 2014) and the Svalbard margin (Jernas et al., 2013; Werner et al., 2013). Sarnthein et al. (2003) postulated pulses of AW inflow to the western Barents Sea shelf at 2.2 and 1.6 cal ka BP. According to Perner et al. (2015), the Neoglacial delivery of chilled AW to the Nordic Seas culminated between 2.3 and 1.4 cal ka BP. These results are in accordance with the timing of major AW inflows revealed by our record.

6 Conclusions

The ST_1.5 multiproxy record revealed that the environmental variability in Storfjorden during the Neoglacial was controlled primarily by the interplay between AW, ArW and sea ice cover variability. The molecular record supports and complements sedimentary and microfossil records, which indicate that major changes in the environmental conditions in Storfjorden occurred at ∼2.7 cal ka BP. The general cooling in the early phase of the Neoglacial initiated conditions for the formation of extensive sea ice cover. The latter part of the
Neoglacial (after \(\sim 2.7 \text{ cal ka BP}\)) was characterized by alternating short-term cooling and warming periods. Warming was associated with pulsed inflows of AW and sea ice melting, which may stimulate phytoplankton blooms and organic matter supply to the bottom. The cold phases were characterized by heavy and densely packed sea ice, resulting in limited productivity.

Moreover, the aDNA diatom record supports the conclusion that primary production took place continuously during the Neoglacial, regardless of the sea ice conditions. The early phase of the Neoglacial was characterized by the presence of diatom taxa associated with sea ice, whereas the present-day diatom assemblage is dominated by *Chaetoceros* spp., a taxa characteristic of open water.

The aDNA record of monothalamous foraminifera is in agreement with the microfossil record and revealed the timing of the major pulses of AW at 2.3 and 1.7 cal ka BP. The AW inflow was marked by an increase in the percentage of sequences of monothalamous taxa associated with the presence of fresh phytodetritus. The monothalamous assemblage during cold intervals was less diverse and was dominated by monothalamous foraminifera known only from environmental sequencing.

**Data availability.** The abundance of fossil foraminifera and the number of foraminiferal and diatom aDNA sequences can be found in the Supplement (Tables S1, S2 and S3).

**Supplement.** The supplement related to this article is available online at: https://doi.org/10.5194/cp-16-487-2020-supplement.

**Author contributions.** MZ and JP designed the study. JP, MLand MZ collected the sediment core. MLand MK performed the sedimentological and micropaleontological analyses. JP performed the molecular analyses and prepared the paper with contributions from all coauthors.

**Competing interests.** The authors declare that they have no conflict of interest.

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